



Review Article

Antifertility activity of medicinal plants

Muhammad Daniyal^{a,*}, Muhammad Akram^b

^a Faculty of Eastern Medicine and Surgery, Hamdard University, Karachi, Pakistan

^b Department of Eastern Medicine and Surgery, Faculty of Medical and Health Sciences, University of Poonch, Rawalakot, Azad Jammu and Kashmir, Pakistan

Received October 1, 2014; accepted November 25, 2014

Abstract

The aim of this review was to provide a comprehensive summary of medicinal plants used as antifertility agents in females throughout the world by various tribes and ethnic groups. We undertook an extensive bibliographic review by analyzing classical text books and peer reviewed papers, and further consulting well accepted worldwide scientific databases. We performed CENTRAL, Embase, and PubMed searches using terms such as “antifertility”, “anti-implantation”, “antiovulation”, and “antispermatogenic” activity of plants. Plants, including their parts and extracts, that have traditionally been used to facilitate antifertility have been considered as antifertility agents. In this paper, various medicinal plants have been reviewed for thorough studies such as *Polygonum hydropiper* Linn, *Citrus limonum*, *Piper nigrum* Linn, *Juniperis communis*, *Achyranthes aspera*, *Azadirachta indica*, *Tinospora cordifolia*, and *Barleria prionitis*. Many of these medicinal plants appear to act through an antizygotic mechanism. This review clearly demonstrates that it is time to expand upon experimental studies to source new potential chemical constituents from medicinal plants; plant extracts and their active constituents should be further investigated for their mechanisms. This review creates a solid foundation upon which to further study the efficacy of plants that are both currently used by women as traditional antifertility medicines, but also could be efficacious as an antifertility agent with additional research and study. Copyright © 2015 Elsevier Taiwan LLC and the Chinese Medical Association. All rights reserved.

Keywords: antifertility agent; efficacy; literature review; medicinal plants

1. Introduction

Antifertility agents are drugs that control fertility¹ and are also called oral contraceptives. These drugs affect and are involved in the menstrual cycle and ovulation in females. Estrogen and progesterone in combined form are given as birth control pills. The antifertility substance is deemed to be active in females when it prevents fertilization, prevents ovulation, implantation, and destroys the zygote or causes abortion. In males, it prevents spermatogenesis, inhibits testosterone, or affects the gonadotrophin of the organs or the mortality of

sperm. Currently, population size is being controlled in many developing countries.² Oxyphenbutazone, indomethacin, and acetyl salicylic acid inhibit prostaglandin formation and manifest antifertility activities in albino male and female rabbits. The reproductive process is affected by indomethacin and oxyphenbutazone in male rabbits. Significant anti-ovulatory activity was exhibited by acetyl salicylic acid 300 mg/kg and indomethacin 3 mg/kg in the rabbits. Additionally, indomethacin exhibited anti-implantation activity. This shows that reproduction in female animals is affected by nonsteroidal anti-inflammatory drugs.³ Antifertility activity in antimalarial drugs has been observed, because amodiaquine blocks ovulation and disrupts the oestrous cycle.⁴ It has also been observed that many plants may have spermicidal activity. Medicinal plants are a great gift of nature as a cure-all for a plethora of human problems. Various institutes in South Asia have long-established traditions of cultivating the faculties of

Conflicts of interest: The authors declare that there are no conflicts of interest related to the subject matter or materials discussed in this article.

* Corresponding author. Dr. Muhammad Daniyal, Faculty of Eastern Medicine and Surgery, Hamdard University, Karachi, Pakistan.

E-mail address: daniyaldani151@yahoo.com (M. Daniyal).

the younger generation in the emerging field of science and technology. Currently, many scientists in that region have an ongoing mandate to educate and train the younger generation to facilitate future innovation and advancement in the field, while also working with venture capital elements and the corporate sector at the same time to optimize their research opportunities. It goes without saying that modern scientific investigation has proven the medicinal value of medicinal plants. Herbal medicines and their derivatives have been incorporated into traditional medicine virtually since the beginning of recorded history. But it is only in recent times that the broader use of medicinal plants is beginning to garner acceptance in the more expansive international domain. There are certain bottlenecks in the process, including but not limited to the lack of quality control and toxicological studies, the imperative to increase product shelf life, and compliance with international regulatory standards that need to be overcome before their full market potential can be realized. Various medicinal plants have antifertility activities (Table 1).^{16,31–50} The Unani system of medicine is an indigenous treatment, very prevalent in South Asia and popular among large populations in India, Pakistan, Bangladesh, and Sri Lanka. More specifically, Unani medicine is part of the South Asian culture where it is practiced among a large segment of the population. Due to its regional popularity, Unani medicine has achieved an

exponential growth over the past 5 decades in South Asia. On a practical basis, Unani medicine is innovative in that its practitioners have accepted and managed to avoid many of the challenges that practicing professionals make, including issues with practice–patient relations, forms of intervention, and disease conceptualization itself. Unani medicine has maintained its popularity in a number of South Asian countries, and is used by > 65% of the total population in Pakistan alone. This knowledge that is presently used to control conception, though still in use among tribal populations, requires further scientific examination, so that the active components present in such plants which are capable of obstructing the reproductive cycle can be properly delineated and utilized. The global herbal medicine sector is expected to reach close to \$115 billion profit by 2015, according to the Global Industry Analyst Market and such growth is fueled by a trend towards herbal and nature-based products based on the presumption these products cause fewer side effects than modern medicines. Approximately 75% of the population in emerging nations receive herbal medical health care, compared with over half of the population in developed nations, particularly for lifestyle-related diseases. The herbal medicine market is also benefiting from a structural change in how it can generate financial gain for different countries and their business sectors, which is why more companies are providing medical coverage

Table 1
List of antifertility medicinal plants.

Plant	Type	Dose/body weight (mg/kg)	Activity	Refs
<i>Cichorium intybus</i>	50% ethanolic extract	50	Anti-implantation	16
<i>Cuscuta reflexa</i>	Ethanolic extract	800	Anti-implantation	16
<i>Rubia cordifolia</i>	Ethanolic extract	250	Anti-implantation	16
<i>Urtica dioica</i>	Ethanolic extract	250	Anti-implantation	16
<i>Abroma augusta</i>	Petroleum ether	50	Anti-implantation	16
<i>Curcuma longa</i>	Petroleum ether	200	Anti-implantation	16
<i>Plumbago rosea</i>	Acetone extract	200	Anti-implantation	16
<i>Aloe barbadensis</i>	Aqueous extract	100	Anti-implantation	16
<i>Abutilon indicum</i>	50% aqueous methanolic extract	500	Anti-implantation	16
<i>Artemisia vulgaris</i>	Methanolic extract	300 and 600	Anti-implantation	31
<i>Striga orobanchioides</i>	Ethanolic extract	200	Anti-implantation	32
<i>Acalypha indica</i> Linn	Ethanolic extract	600	Estrogenic activity	33
<i>Enicostemma axillare</i>	Ethanolic extract	375 and 750	Antispermato-genic	34
<i>Mondia whitei</i> Linn	Ethanolic extract	400	Antispermato-genic	35
<i>Moringa oleifera</i>	90% ethanol extract	175	Abortifacient	36
<i>Curcuma longa</i> Linn	70% alcoholic extract	500	Antispermato-genic	37
<i>Abrus precatorius</i> Linn	70% methanolic extract	20 and 40	Antifertility	38
<i>Aegle marmelos</i>	50% ethanolic extract	100, 200, and 300	Antifertility effect	39
<i>Albizia lebeck</i>	Methanolic extract	50, 100, and 200	Antifertility effect	40
<i>Bacopa monnieri</i>	Dry powder	250	Antispermato-genic	41
<i>Cannabis sativa</i>	Alcoholic extract	20	Antispermato-genic	42
<i>Dendrophthoe falcata</i>	70% methanolic extract	100	Antispermato-genic	43
<i>Fadogia agrestis</i>	Aqueous extract	18, 50, and 100	Adverse effects on male rat testicular function	44
<i>Juniperus phoenicea</i>	Ethanolic extract	Intraperitoneal injections of 400	Antifertility activity	45
<i>Leptadenia hastata</i>	Aqueous extract	100, 200, 400, and 800	Antispermato-genic activity	46
<i>Ocimum sanctum</i>	Benzene extract	300	Antifertility property	47
<i>Quassia amara</i>	Chloroform extracts	Single daily intramuscular injections of the extract for 15 d	Antifertility effect	48
<i>Syzygium aromaticum</i>	Hexane extract	15, 30, and 60	Degenerative changes in the seminiferous tubules	49
<i>Terminalia bellirica</i>	Alcoholic extracts	50 mg/d	Antifertility effect	50

for herbal medical care. One major obstacle to industry growth is the lack of herbal medicine education institutions and the relatively moderate amount of resources devoted to research and development. The degree of risk associated with alternative therapies is perceived as a sort of elevated or higher education of herbal medicine institutions, which is not yet fully understood by many medical professionals in the mainstream medical community. The antifertility properties of medicinal plants and their utility in human diseases for which plants have now achieved a global reputation have been experimentally confirmed around the globe over a substantial number of years. Medicinal plants have been studied for their antifertility activity, for example, various parts of different plants contain estrogen and have their own effects on different life processes. Phytoestrogens are novel agents found in a variety of plants. They have noxious effects leading to impaired fertility in domestic animals by disturbing the normal gestation process. Day by day, the world population inevitably increases due to a global lack of family planning and support. There are various methods of family planning, though all of these methods generally have some side effects. Some methods used for antifertility permanently arrest spermatogenesis and cause irreversible sterility. The extent of fertility in females and males are also being controlled by vaccines. The active mechanism in such a vaccine is typically the production of antibodies against human chorionic gonadotropin, though this vaccine causes sterility in male animals.⁵ Additionally, the progression of spermatogenesis is suppressed in males by antifertility drugs. At this time, various drugs such as triptolide, tamoxifen, gossypol, and testosterone are being used as antifertility agents.⁶ Clinicians working in Europe, North America, and other industrialized countries are taking an interest in research dealing with ethnopharmacology. Some of the medicinal plants discussed appear promising and may possibly lead to the development of products equally as effective as their corresponding existing drug. Furthermore, in an allopathic system, an effective abortifacient (misoprostol) can be administered orally or sublingually, and is extremely cheap but does have side effects. Consequently, due to existing side effects of allopathic medicines, people are afraid of using these medicines. A review of the literature indicated the potential benefits of the use of a number of plants/preparations for fertility regulation. Some local contraceptive agents have also been described in traditional medicine. An attempt has been made to document medicinal plants that are usually prescribed as antifertility agents or have been tested for their activity *in vitro* or *in vivo*.

2. Methodology

A bibliographic investigation was carried out by analyzing classical text and reference books, articles, and peer-reviewed papers, as well as a thorough consultation of worldwide accepted scientific databases. We performed CENTRAL, Embase, and PubMed searches using terms such as “antifertility”, “anti-implantation”, “antiovarulation”, and “antispermato-genic” activity of plants.

3. Medicinal plants used as antifertility agents

3.1. Antiovarulation activity

Polygonum hydropiper Linn belongs to the family Polygonaceae, which is in part valued for its roots and leaves and includes such active ingredients as formic acid, acetic acid, beldianic acid, tannin, essential oil, and oxymethyl-anthraquinones. It is used in cases involving diarrhea, skin problems, hemorrhoids, and dyspepsia. Biologically, these ingredients can have antioxidant, antimicrobial, anti-inflammatory, and antifertility effects in humans. In one study, Kapoor et al⁷ have reported on the antiovarulatory activity in this plant. Their study using three types of extracts (petroleum, aqueous, and alcohol) was conducted to investigate the antifertility activity of this particular plant. Antifertility activity was observed in rabbits with copper-induced ovulation. Petroleum ether extract of the roots of *Polygonum hydropiper* was found effective in inhibiting ovulation in 60% of the animals. Additionally, $\leq 40\%$ inhibition was observed by all other extracts.

3.2. Anti-implantation activity

Calotropis gigantea Linn belongs to the family Asclepiadaceae. Here, the plant part primarily used in antifertility treatments is the leaves, which operate within the active principles of glycosides and cardenolides. It is used for cases of colic, flatulence, asthma, cough, and whooping cough. The biological activities associated with this plant facilitate wound healing, have anticancer and hypoglycemic effects, as well as antifertility properties. Antifertility activity arising from *Calotropis gigantea* root has been reported in rats.⁸ In one study, ethanolic extract was administered to albino rats to investigate its antifertility effect; this extract showed strong anti-implantation activity at the dose level of 250 mg/kg.⁹ Another plant, *Hibiscus rosa-sinensis* belongs to the family Malvaceae, the active parts of which are believed to be its flowers. Its active principles include quercetin-7-O-galactoside, polyphenolic compounds, kaempferol, and scutellarein and it is used to treat bacterial infection, hyperlipidemia, and depression. Its asserted biological activity is antibacterial, antioxidant, hypolipidemic, and antispermato-genic. In another study, Neeru and Sharma¹⁰ investigated the antifertility and estrogenic activity of *Hibiscus rosa-sinensis* Linn by utilizing an ethanolic extract of this plant. The plant extract exhibited strong anti-implantation and uterotrophic activity (inhibition was 100%) at a dose level of 400 mg/kg.¹⁰ Women and local physicians use *Hibiscus rosa-sinensis* for its benzene extract-related antifertility activity, which was observed to cause an 80% decrease in implantation site activity on the 10th day of pregnancy.¹¹ The extract from another plant, *Butea monosperma*, was fed to female mice and decreased fertility was observed.¹² Khanna et al¹³ reported that *Butea monosperma* has antifertility activity that was revealed in its hot alcohol extract. There were no implantation sites found in rats that were fed *Butea monosperma* extract at a dose of 300 mg/kg.¹³

From another plant, the aqueous extract of *Ocimum sanctum* has anti-implantation activity that was revealed in >50% of the subject rats.¹⁴ Additionally, *Striga orobanchioides* has anti-implantation activity that is also observed in rats. It was noted that epithelial cell height, thickness of endometrium, increase in diameter, and increase in uterine weight were observed in rats after administration of two flavones, luteolin and apigenin, from *Striga orobanchioides* as compared to control rats.¹⁵ Maurya et al¹⁶ reported that the ethanolic extract of seeds of *Ricinus communis* (80 mg/kg), fruits of *Punica granatum* (1.82 mg/kg), roots of *Calotropis procera* (30 mg/kg), roots of *Polygonum hydropiper* (150 mg/kg), leaves of *Mentha arvensis* (100 mg/kg), leaves of *Lawsonia inermis* (134 mg/kg), seeds of *Juniperus communis* (200 mg/kg), roots of *Hagenia abyssinica* (120 mg/kg), seeds of *Crotalaria juncea* (300 mg/kg), and roots of *Cicer arietinum* (1900 mg/kg) all have anti-implantation activity. *Citrus limonum* belongs to the family Rutaceae, where its ripe fruit, root, and leaves are all used to promote antifertility outcomes. Its active principles include limonoid glycosides, limonene, geraniol, neral, ichangin 4-beta-glucopyranoside, nomilinic acid, and 4-beta-glucopyranoside. The derivatives arising from this plant are used to treat scurvy, arthritis, and uterine hemorrhage. Its biological activity is stomachic, carminative, and anthelmintic. Kulkarni et al¹⁷ reported in their study about the antifertility effect of lemon seeds (*Citrus limonum*) in female albino mice. This study was performed using female albino mice and petroleum ether, alcohol, and aqueous extract were administered to gauge antifertility activity. The extracts were administered orally for 7 days after insemination (i.e., post-ovulatory test) and 4% gum acacia was given to mice in the control group. On the 10th day of pregnancy, animals were investigated for implantation sites. It was determined that the activity of alcoholic extract exceeded that of both the petroleum ether and aqueous extracts. Alcohol extract was fractionated and these fractions were further evaluated to assess their antifertility properties. Overall the most significant activity was observed by fraction of ethyl acetate. Furthermore, alcoholic extract and ethyl acetate fraction were investigated to more accurately ascertain their possible mechanism of action. Ultimately, they both were confirmed to have antizygotic agent characteristics. When the test drug was withdrawn, there was a complete restoration of fertility. This study indicated that ethyl acetate fraction of alcoholic extract of lemon seeds exhibited reversible antifertility activity in female mice due to its antizygotic activity.¹⁷ *Achyranthes aspera* belongs to the family Amaranthaceae, where those parts used consist of its leaves and seeds. It has been demonstrated that its active principles include calcium, potassium, saponins, ecdysterone, achyranthine, and inokosterone. Currently, it is used to treat asthma and cough. Previously, Shibeshi et al¹⁸ reported on the antifertility effect of *Achyranthes aspera* Linn, indicating that its biological activity is antimicrobial, hypolipidemic, and antifertility. In one study, the antifertility activity in fertile female albino rats of *Achyranthes aspera* Linn was investigated. The orally administered dosage of ethanolic extract of this plant was 200 mg/kg, with a treatment duration of 1–7

days of pregnancy. Most notably, an 83.3% anti-implantation activity was exhibited by the ethanol extract of plant at a dose of 200 mg/kg body weight, anti-implantation and abortifacient activity was 100%.¹⁹

4. Antispermatic activity

Plumbago zeylanica belongs to the family Plumbaginaceae and its antifertility components include roots and leaves. Its active principles are plumbagin, isoshinanolone, trans-cinnamic acid, vanillic acid, beta-sitosterol, 4-hydroxybenzaldehyde, and plumbagic acid and it is used to treat piles, leukoderma, and other skin diseases. It appears to foster diverse biological activities including antihelicobacter pylori, antidiabetic, antioxidant, and antifertility. An earlier rat study was undertaken using the plant's ethanol extract. When the applied extract dosage was 159 mg/kg, seminiferous tubules diameters were decreased and spermatocytes and spermatids production was reduced. Furthermore, a reduction in immature and mature Leydig cells occurred and degenerating cells were significantly increased. Lastly, the testicular cell population was decreased. Overall, this study showed palpable plant based antifertility activity.²⁰ *Piper nigrum* Linn belongs to the family Piperaceae, where the parts used are fruits. The active principles are thujon, piperettine, piperolin A, piperolin B, terpene, volatile oil, starch, piperine, calcium, phosphorus, iron, thiamine, riboflavin, nicotinic acid, vitamin C, carotene, and piperidine. Derivatives of this plant are used in cases of arthritis, inflammation, asthma, dyspepsia, flatulence, nervous debility, liver affections, urine retention, amenorrhea, sexual weakness, enteritis, intermittent fever, toothache, piles, leukoderma, leprosy, neuralgia, constipation, abdominal pain, paralysis, general weakness, fever, and baldness. Its biological activity has been observed to be abortifacient, antipyretic, and antiasthmatic. In an article by Mishra and Singh²¹ in 2009, the authors reported on the antispermatic and antifertility effects of the fruits of *Piper nigrum* Linn in mice. In this study, the drug was administered orally at a dose of 25 mg/kg/d and 100 mg/kg/d, for 20 days and 90 days and the efficacy of this plant was investigated, associated with their impact on the reproductive organs of male mice. The end points for assessments were fertility indices, organ weight, fructose content, sialic acid, histopathology, and sperm parameters. There were significant degenerative changes after 90 days of treatment with *Piper nigrum* and sperm parameters were also affected.²¹ *Azadirachta indica* belongs to family Meliaceae and those parts suspected of having antifertility properties are the seeds and leaves. The active principles are margosic acid, azadirachtin, polysaccharides, nimbine, nimbinate, nimbidol, oil, and nimbidin. Extracts from this plant are sometimes used to treat skin disorders and the biological activities are hypoglycemic, antioxidant, antiviral, and spermicidal. In one study performed on male rats, *Azadirachta indica* extract was investigated for antifertility activity. The study duration was 30 days with a drug dosage of 10 mg. A 50% ethanol extract of *Azadirachta indica* given subcutaneously, which when administered for 30 days arrested spermatogenesis.

Furthermore, the size of epididymides, Leyding cell nuclei, and seminiferous tubules also were significantly reduced and the reduction in sperm motility was significant as well. It would appear that the arrest of spermatogenesis and androgen depletion was due to reduced testicular and accessory organ protein, sialic acid, and vesicular fructose content.²² *Striga lutea* belongs to the family Orobanchaceae, and all parts of the plant have antifertility activity. Here, active principles are acacetin, luteolin, and flavones. It is used as an anti-implantation agent with its principle biological activity being its antifertility activity. Hiremath et al²³ undertook a study which reported on the antifertility capacity of this plant. Chloroform and petroleum ether extracts were used as part of the investigation. The results showed significant antifertility activity of *Striga lutea* in mice. An effective dose of chloroform extract was 50 mg/kg and petroleum ether extract was effective at 100 mg/kg. Antifertility activity was confirmed upon histological examination of the subject's uterus and ovary.²³ *Barleria prionitis* belongs to the family Acanthaceae. Parts of the plant that were used included the roots, leaves, and stem. Active principles are iridoid glycosides, barlerin, and verbascoside. It is used by people to address body ache, tooth ache, and bleeding gums. The suspected biological activity is hepatoprotective, antifertility, anti-inflammatory, immunomodulant, and antidiabetic. One recent antifertility study of the root extract of *Barleria prionitis* Linn in male albino rats has been reported. *Barleria prionitis* L. was given orally to male rats (100 mg/d). The duration of study was 60 days and the extract reduced the fertility of male rats by 100%.²⁴ *Tinospora cordifolia* belongs to the family Menispermaceae, and the plant stem was alleged to be the active part. Active principles are berberine, gilion, gilossterol, columbin, chasanthin, palmarin, tinosporin, alkaloid, tinosporic acid, and tinosporol. It is used to treat tuberculosis, fever, and wounds. The asserted biological activity is antifertility, antioxidant, hypoglycemic, immunomodulant, and cardioprotective. In that study, the antifertility effect of *Tinospora cordifolia* (Willd.) stem extract in male rats has been reported. Overall, 70% methanolic extract of *Tinospora cordifolia* stem was given orally to male rats at the dose level of 100 mg/d. The study's duration was 60 days during which time there was 100% suppression of spermatogenesis and reduction of male fertility.²⁵

4.1. Effect on hormones

Martynia annua Linn belongs to the family Martyniaceae and its fruits and seeds are those parts which may have antifertility properties. Its active principle is luteolin and is used to treat skin infection and eczema. With this plant, the biological activity is wound healing and antifertility. In a study, 50% ethanol extract of *Martynia annua* was used to investigate its effects on male rats. There were four groups of rats and each group comprised of five animals. The first group served as a control group. An extract of 50 mg/kg was given to the second group, and 100 mg/kg of extract was administered to the third group. Lastly, a 200 mg/kg extract was administered to the fourth group. This extract was given daily for 60 days. The

weights of testes, ventricle vesicles, epididymides, and seminal vesicle were significantly reduced accompanied by a reduced testicular sperm count. The severity of seminiferous epithelium lesions was dependent on the dose of the extract. Also, seminiferous tubules became reduced in size. At the secondary stage, spermatogenesis was arrested and pachytene spermatocytes degeneration was observed. Immature germ cells were damaged and sloughed. Atrophy of Leydig cells was observed. There were no morphological changes in Sertoli cells but luteinizing hormone and testosterone were reduced significantly. Concentration of follicle-stimulating hormone was not altered significantly. There were no changes in the parameters of hematology. This study indicates that extracts produce dose-related effects on male reproduction without altering the metabolism of the general body.²⁶ *Juniperis communis* belongs to the family Cupressaceae which has potential antibacterial properties contained in the roots. Its active principles are glycosides, formic acid, acetic acid, diterpine acid, resin, volatile oil, juniperin, vitamin C, and tannins. This plant is used in cases of urine retention, bladder problems, leucorrhea, gonorrhoea, vaginal infections, scarlet fever, smallpox, and cholera. Primarily, its biological activity is diuretic and a digestive tonic. In an earlier study, Sandhya et al²⁷ reported on the antiprostaglandin activity of this plant which accounts for its antifertility effect. Additionally, *Juniperus communis* was investigated for its antifertility properties. Its several activities such as those promoting estrogenic, antiestrogenic, progestagenic, and anti-progestagenic mechanisms were investigated. Ultimately, its activity as an antiprogestational was significant in that it is attributed to its antifertility effect. *Andrographis paniculata* also arrests the spermatogenesis in male albino rats which is due to the andrographolide present in *Andrographis paniculata*.²⁷ Alpha-chlorohydrin is a male contraceptive. Oral use of alpha-chlorohydrin causes antifertility and reverses after withdrawal of alpha-chlorohydrin. Nephrotoxic and neurotoxic affects has been observed at higher doses of alpha-chlorohydrin.²⁸ Additionally, a reduction in weight of the seminal vesicle, epididymis, and testis and an increase in the anterior pituitary gland was observed in male rats given *Quassia amara* extract. The plant extracts of *Quassia amara* reduce follicle-stimulating hormones, luteinizing hormones, serum levels of testosterone, and epididymal sperm counts.²⁹

4.2. Abortifacient activity

Spondias mombin belongs to the family Anacardiaceae where its leaves are the suspected antifertility part of the plant. Active principles are flavonoids, quercetin, ellagic acid, and rutin. It is used in the treatment of insomnia, epilepsy, and psychosis and is endowed with biological activity including antioxidant and antibacterial. In an earlier study, authors investigated the *in vivo* efficacy of this plant extract on the reproductive performance of female rats. The plant was evaluated for anticonceptive and abortifacient activity. Ovariectomized rats were used to test estrogenic activity. Acute toxicity activity of this plant was also assessed and it was found to be relatively nontoxic. Although, this plant exhibited

anticonceptive activity, it had no abortifacient activity evident from the number of pregnant animals at the end of the third trimester of pregnancy. Additionally, estrogenic activity was not exhibited by the extract. However, the study indicated that aqueous ethanol extract of the plant has significant anti-conceptive activity.³⁰

In conclusion, this review aims to promote the use of plants and their extracts arising from their antifertility activities. Already, medicinal plants have a proven efficacy as antifertility agents. The mechanism of action of many herbs has been identified. Further research is required to better ascertain the bioactivity of other compounds in crude extracts and to exploit their activity as antifertility agents. As such, new and efficacious drugs can be developed by way of comprehensive investigation of the bioactivity of various compounds.

References

- Kumar D, Kumar A, Prakash O. Potential antifertility agents from plants: a comprehensive review. *J Ethnopharmacol* 2012;**140**:1–32.
- Pei X, Nai W. Can ethnopharmacology contribute to the development of antifertility drugs? *J Ethnopharmacol* 1991;**32**:167–77.
- Yegnanarayan R, Joglekar G. Antifertility effect of non-steroidal anti-inflammatory drugs. *Jpn J Pharmacol* 1978;**28**:909–17.
- Gbotolorun S, Osinubi A, Oremosu A, Noronha C. The antifertility effect of amodiaquine hydrochloride. *Nig Q J Hosp Med* 2011;**21**:271–5.
- Talwar G, Raghupathy R. Antifertility vaccines. *J Vacc* 1989;**7**:97–101.
- Zha S, Zha J, Huang Y. Male antifertility drugs and cell apoptosis. *National J Androl* 2008;**14**:75–8.
- Kapoor M, Garg SK, Mathur V. Anthiovoluntary activity of five indigenous plants in rabbits. *Indian J Med Res* 1974;**62**:1225–7.
- Srivastava SR, Keshri G, Bhargavan B. Pregnancy interceptive activity of the roots of *Calotropis gigantea* Linn. in rats. *J Contracep* 2007;**4**:318–22.
- Kamath JV, Rana A. Preliminary study on antifertility activity of *Calotropis procera* roots in female rats. *Fitoterapia* 2001;**73**:111–5.
- Neeru V, Sharma S. Post-coital antifertility activity of *Hibiscus rosasinensis* Linn. roots. *Evid Based Complement Alternat Med* 2008;**5**:91–4.
- Kirtikar K, Basu B. Indian medicinal plants. *Periodical Exp* 1935;**2**:848–9.
- Dreisbach R. Effect of drugs on reproduction in mice. *Indian J Physiol Pharmacol* 1963;**7**:63–71.
- Khanna U, Chaudhury R. Antifertility screening of plants, 3333 Part I. Investigation on *Butea monosperma* Lam. Kuntz. *Indian J Med Res* 1968;**56**:1577–80.
- Vohora S, Garg S, Chaudhury R. Antifertility Screening of Plants. III. Effects of six indigenous plants on early pregnancy in albino rats. *Indian J Med Res* 1969;**51**:893.
- Hiremath S, Badami S, Hunasagatta S, Patil S. Antifertility and hormonal properties of flavones of *Striga orobanchioides*. *Europ J Pharmacol* 2000;**391**:193–7.
- Maurya R, Srivastava S, Kulshreshta D. Traditional remedies for fertility regulation. *Curr Med Chem* 2004;**11**:1431–50.
- Kulkarni T, Kothekar MA, Mateenuddin M. Study of antifertility effect of lemon seeds (*Citrus limonum*) in female albino mice. *Ind J Physiol Pharmacol* 2005;**49**:305–12.
- Shibeshi W, Eyasu M, Legesse Z. Effect of *Achyranthes aspera* L. on fetal abortion, uterine and pituitary weights, serum lipids and hormones. *Afr Health Sci* 2006;**6**:108–12.
- Vasudeva N, Sharma S. Post-coital antifertility activity of *Achyranthes aspera* Linn. root. *J Ethnopharmacol* 2006;**107**:179–81.
- Parohit A, Surendra K, Keshav B. Contraceptive efficacy of *Plumbago zeylanica* root extract (50% ETOH) in male albino rats with special emphasis on testicular cell population dynamics. *Anc Sci Life* 2008;**27**:31–5.
- Mishra RK, Singh S. Antispermato-genic and antifertility effects of fruits of *Piper nigrum* L. in mice. *Ind J Exp Biol* 2009;**47**:706–14.
- Parohit A. Antifertility efficacy of neem bark (*Azadirachta indica* A.juss.) in male rats. *Anc Sci Life* 1999;**19**:21–4.
- Hiremath SP, Rao SH, Jain P. Antifertility activity of striga lutea-Part I. *Indian J Physiol Pharmacol* 1990;**34**:23–5.
- Gupta RS, Kumar P, Dixit V. Antifertility studies of the root extract of the *Barleria prionitis* Linn in male albino rats with special reference to testicular cell population dynamics. *J Ethnopharmacol* 2000;**70**:111–7.
- Gupta RS, Sharma A. Antifertility effect of *Tinospora cordifolia* (Willd.) stem extract in male rats. *Indian J Exp Biol* 2003;**41**:885–9.
- Mali PC, Ansari AS, Chaturvedi M. Antifertility effect of chronically administered *Martynia annua* root extract on male rats. *J Ethnopharmacol* 2002;**82**:61–7.
- Sandhya P, Tewari R, Prakash A. Hormonal properties of ethanolic extract of *Juniperus communis* Linn. *Anc Sci Life* 1990;**10**:106–13.
- Jones A. Antifertility actions of alpha-chlorohydrin in the male. *Aust J Biol Sci* 1983;**36**:333–50.
- Y1 Raji, Bolarinwa AF. Antifertility activity of *Quassia amara* in male rats - *in vivo* study. *Life Sci* 1997;**61**:1067–74.
- Chukwuka N, Uchendu T. Antifertility activity of aqueous ethanolic leaf extract of *Spondias mombin* (Anacardiaceae) in rats. *Afr Health Sci* 2008;**8**:163–7.
- Shaik A, Kanhere R, Cuddapah R, Nelson K, Vara P, Sibyala S. Antifertility activity of *Artemisia vulgaris* leaves on female Wistar rats. *Chin J Nat Med* 2014;**12**:180–5.
- Hiremath S, Badami S, Swamy K, Patil S, Londonkar R. Antifertility activity of *Striga orobanchioides*. *Biol Pharm Bull* 1994;**17**:1029–31.
- Hiremath S, Rudresh K, Badami S, Patil S, Patil S. Post-coital antifertility activity of *Acalypha indica* L. *J Ethnopharmacol* 1999;**67**:253–8.
- Dhanapal R, Ratna J, Gupta M, Sarathchandran I. Preliminary study on antifertility activity of *Encicostemma axillare* leaves and *Urena lobata* root used in Indian traditional folk medicine. *Asian Pac J Trop Med* 2012;**5**:616–22.
- Watcho P, Kamtchouing P, Sokeng S, Moundipa P, Tantchou J. Reversible antispermato-genic and antifertility activities of *Mondia whitei* L. in male albino rat. *Phytother Res* 2001;**15**:26–9.
- Nath D, Sethi N. Commonly used Indian abortitacient plants with special reference to their teratologic effect in rats. *J Ethanopharmacol* 1992;**36**:147–54.
- Purohit A, Bhagat M. Contraceptive effect of *Curcuma longa* (L.) in male albino rat. *Asian J Androl* 2004;**6**:71–4.
- Bhatt N, Chawla SL, Rao MV. Contraception evaluation of seed extracts of *Abrus precatorius* L. in male albino rats (*Mus musculus*). *J Herb Med Toxicol* 2007;**1**:45–8.
- Chauhan A, Agarwal M, Kushwaha S, Mutreja A. Antifertility studies of *Aegle marmelos* Corr., an Indian medicinal plant on male albino rats. *Egypt J Biol* 2008;**10**:28–35.
- Gupta RS, Kachhawa JB, Chaudhary R. Antifertility effects of methanolic pod extract of *Albizia lebbbeck* (L.) Benth in male rats. *Asian J Androl* 2004;**6**:155–9.
- Singh A, Singh SK. Evaluation of antifertility potential of Brahmi in male mouse. *J Contracep* 2009;**79**:71–9.
- Sailani MR, Moeini H. Effect of *Ruta graveolens* and *Cannabis sativa* alcoholic extract on spermatogenesis in the adult Wistar male rats. *Indian J Urol* 2007;**23**:257–60.
- Gupta RS, Kachhawa JB, Sharma A. Effect of methanolic extract of *Dendrophthoe falcata* stem on reproductive function of male albino rats. *J Herb Pharmacother* 2007;**7**:1–13.
- Yakubu MT, Akanji MA, Oladiji T. Effects of oral administration of aqueous extract of *Fadogia agrestis* (Schweinf. Ex Hiern) stem on some testicular function indices of male rats. *J Ethnopharmacol* 2008;**115**:288–92.
- Shkukani HG, Salhab AS, Disi AM, Shomaf MS, Al Quadan F. Antifertility effect of ethanolic extract of *Juniperus phoenicea* (L.) in male albino rats. *J Herb Pharmacother* 2008;**7**:179–89.
- Bayala B, Telefo PB, Bassole IHN, Tamboura HH, Belemtougri RG, Sawadogo L, et al. Anti-spermato-genic activity of *Leptadenia hastata*

- (Pers.) Decne leaf stems aqueous extracts in male wistar rats. *J Pharmacol Toxicol* 2011;**6**:391–9.
47. Reghunandan R, Sood S, Reghunandan V, Mehta RM, Singh G. Effect of *Ocimum sanctum* Linn (tulsi) extract on testicular function. *Indian J Med Sci* 1995;**49**:83–7.
 48. Parveen S, Das S, Kundra CP, Pereira BM. A comprehensive evaluation of the reproductive toxicity of *Quassia amara* in male rats. *Reprod Toxicol* 2003;**17**:45–50.
 49. Owoyele BV, Soladoye A. Analgesic and anti-inflammatory activity of ethanolic extract of *Chromolaena odorata* leaves. *Recent Progr Med Plants* 2007;**18**:397–406.
 50. Verma PK, Sharma A, Mathur A, Sharma P, Gupta RS, Joshi SC, et al. Effect of *Sarcostemma acidum* stem extract on spermatogenesis in male albino rats. *Asian J Androl* 2002;**4**:43–7.