

Heparanase-1 facilitates bone remodeling on bone defect animal model with optimal dosage and treatment duration

Jir-You Wang^{a,b,c}, Cheng-Fong Chen^{a,b,*}

^aDepartment of Orthopaedics, Taipei Veterans General Hospital, Taipei, Taiwan, ROC; ^bDepartment of Orthopaedics, Therapeutical and Research Center of Musculoskeletal Tumor, Taipei Veterans General Hospital, Taipei, Taiwan, ROC; ^cDepartment of Traditional Medicine, School of Medicine, National Yang-Ming University, Taipei, Taiwan, ROC

Bone is a well model to investigate regeneration or tissue engineering. Bone processes regeneration to remodel, while bone defect occurs, such as trauma, bone resection due to tumor surgery, pathological bone loss, or osteoporosis.¹ Under control by numerous and critical factors, bone remodeling procedure is affected by disease risks to nonunion, such as diabetes, obesity, or inflammatory arthritis. Bone defect is a challenge for orthopedic surgeons due to the limitation of bone grafting treatments, such as inadequate amount of bone materials or lacking of donor.² Tissue engineering with the development of various types of biomaterial scaffolds is a potential way for assist bone regeneration.

The bone defect animal model is well established for preclinical testing of various biomaterials, different compounds, or methods that improve bone remodeling^{1,3} before using on human beings. The suitable animal model should be established according to the study design and the bearing capacity of the treatment bone during the experiments.⁴ Different from tumor model that should be select suitable species for tumor cell types and inject into individual site for each tumor types,⁵ the selection criteria of bone defect model depend on animal availability, acceptability to society, tolerance to captivity, and the physiological and pathophysiological analogies in comparison to humans.⁶

The most commonly used animal models for bone defect are rabbit, rodent, or pigs according to the availability. The rodent model is the most commonly used model, and it is easy to handle. However, the limitation is the small size and thin long bones that hard to create bone defect on long bone with fixation compare to other large animals. Most bone defect models on rodent create the defect location on skull instead of long bones.^{7,8} Chiu et al⁹ created bone defect on mouse distal femur with carefully take over the risk of leg fixation and broken from the defect site was a well-done mouse bone defect model for further experiments. With the assistance of image evidence, the remodeling on bone regeneration can be detected and evaluated under different treatments.

Received June 8, 2020; accepted June 9, 2020.

doi: 10.1097/JCMA.00000000000391.

Heparanase is an endo- β -D-glucuronidase that first isolated from human placenta and also from platelet. This enzyme is specifically cleave heparan sulfate proteoglycans that remodel extracellular matrix.¹⁰⁻¹² Being noticed to increase in various types of cancers, the expression of heparanase-1 may relate to the malignancy and poor prognosis of tumor and plays important role on tumor angiogenesis and metastasis.¹³

In osteogenesis, the overexpression of heparanase-1 was reported to benefit for bone healing by increasing bone mass and enhancing blood flow or angiogenesis in bone defect site.¹² However, the opposite reports were announced that heparanase showed suppression effect of osteogenesis.¹⁴

In the March issue of the Journal, Chiu et al⁹ established a well bone defect model on mice for bone healing investigation and tried different doses of heparanase recombinant protein for osteogenesis evaluation. The result showed that low dose enhanced while high dose suppressed the remodeling of bone defect. The results indicated that the optimal dosage, route, and application period were required to prevent the occurrence of potential adverse effects in the event of overdose.⁹ The findings of this study gave a new insight that though heparanase has the effect to improve bone regeneration, however, the critical dosage is important to be considered for optimal treatment. The further mechanism and potential signaling pathway needs more investigations about the effect of heparanase on osteogenesis.

REFERENCES

- Liu M, Lv Y. Reconstructing bone with natural bone graft: a review of in vivo studies in bone defect animal model. *Nanomaterials (Basel)* 2018;8:999.
- Han Z, Bhavsar M, Leppik L, Oliveira KMC, Barker JH. Histological scoring method to assess bone healing in critical size bone defect models. *Tissue Eng Part C Methods* 2018;24:272–9.
- 3. El-Rashidy AA, Roether JA, Harhaus L, Kneser U, Boccaccini AR. Regenerating bone with bioactive glass scaffolds: a review of in vivo studies in bone defect models. *Acta Biomater* 2017;62:1–28.
- McGovern JA, Griffin M, Hutmacher DW. Animal models for bone tissue engineering and modelling disease. *Dis Model Mech* 2018;11:dmm033084.
- Wang JY, Wu PK, Chen PC, Lee CW, Chen WM, Hung SC. Generation of osteosarcomas from a combination of Rb silencing and c-Myc overexpression in human mesenchymal stem cells. *Stem Cells Transl Med* 2017;6:512–26.
- Li Y, Chen SK, Li L, Qin L, Wang XL, Lai YX. Bone defect animal models for testing efficacy of bone substitute biomaterials. *J Orthop Translat* 2015;3:95–104.

^{*}Address correspondence. Dr. Cheng-Fong Chen, Department of Orthopaedics & Traumatology, Taipei Veterans General Hospital, 201, Section 2, Shi-Pai Road, Taipei 112, Taiwan, ROC. E-mail address: cf_chen@vghtpe.gov.tw (C.-F. Chen). Conflicts of interest: The authors declare that they have no conflicts of interest related to the subject matter or materials discussed in this article. Journal of Chinese Medical Association. (2020) 83: 799-800.

Copyright © 2020, the Chinese Medical Association. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/ by-nc-nd/4.0/)

- 7. Sasaki T, Niizuma K, Kanoke A, Matsui K, Ogita S, Rashad S, et al. Octacalcium phosphate collagen composite (OCP/Col) enhance bone regeneration in a rat model of skull defect with dural defect. *Heliyon* 2020;6:e03347.
- Guo K, Liu ZL, Wang WC, Xu WF, Yu SQ, Zhang SY. Chitosan oligosaccharide inhibits skull resorption induced by lipopolysaccharides in mice. *BMC Oral Health* 2019;19:263.
- 9. Chiu PY, HuangFu WC, Liu IH, Chang YP. Topical application of Heparanase-1 facilitates bone remodeling during the healing of bone defects in a mouse model. *J Chin Med Assoc* 2020;83:272–9.
- Vlodavsky I, Gross-Cohen M, Weissmann M, Ilan N, Sanderson RD. Opposing functions of heparanase-1 and heparanase-2 in cancer progression. *Trends Biochem Sci* 2018;43:18–31.
- 11. Xu X, Ding J, Ding H, Shen J, Gattuso P, Prinz RA, et al. Immunohistochemical detection of heparanase-1 expression in eutopic and ectopic endometrium from women with endometriosis. *Fertil Steril* 2007;88:1304–10.
- Rodrigues LM, Theodoro TR, Matos LL, Mader AM, Milani C, Pinhal MA. Heparanase isoform expression and extracellular matrix remodeling in intervertebral disc degenerative disease. *Clinics (Sao Paulo)* 2011;66:903–9.
- Barbosa GO, Bruni-Cardoso A, da Silva Pinhal MA, Augusto TM, Carvalho HF. Heparanase-1 activity and the early postnatal prostate development. *Dev Dyn* 2019;248:211–20.
- 14. Ruan J, Trotter TN, Nan L, Luo R, Javed A, Sanderson RD, et al. Heparanase inhibits osteoblastogenesis and shifts bone marrow progenitor cell fate in myeloma bone disease. *Bone* 2013;57:10–7.